

PE 6 - Crystal Structure and Enzymatic Properties of a Bacterial Family 19 Chitinase Reveal Differences with Plant Enzymes

I.A. Hoell⁽¹⁾, B. Dalhus⁽²⁾, E.B. Heggset⁽¹⁾, S.I. Aspmo⁽¹⁾, V.G.H. Eijsink⁽¹⁾

⁽¹⁾Department of Chemistry, Biotechnology and Food Science, Norwegian University of Life Sciences, P.O. Box 5003, N-1432 Ås, Norway - ⁽²⁾Institute of Medical Microbiology, Section for Molecular Biology, National University Hospital, N-0027 Oslo, Norway

We describe the cloning, over-expression, purification, characterization and crystal structure of ChiG, a single domain family 19 chitinase from the Gram-positive bacterium *Streptomyces coelicolor* A3(2). Family 19 chitinases are commonly found in many plants, but were only recently discovered in bacteria. Studies of the enzymatic properties of ChiG revealed that the enzyme is active towards a variety of soluble chitinous substrates at rates similar to those observed for other chitinases, but has a very limited ability to degrade crystalline chitin. The crystal structures of ChiG and a related *Streptomyces* chitinase, ChiC, showed that bacterial family 19 chitinases lack several loops that extend the substrate-binding grooves in family 19 chitinases from plants. In accordance with these structural features, detailed analysis of the degradation of chito-oligosaccharides by ChiG showed that the enzyme has only four subsites (as opposed to six for plant enzymes). This finding, in combination with the fact that ChiG has a very open active site, can be an explanation for the low activity towards crystalline substrates. Previous studies have shown that chitinases with high activity against crystalline chitin, such as *Bacillus circulans* chitinase A1 and *Serratia marcescens* chitinase A, have deep substrate-binding grooves. In addition, these grooves contain linearly aligned aromatic residues that may play an important role in guiding a chitin chain from the crystalline chitin surface to the catalytic center. Such a stretch of aromatic residues is lacking in ChiG. The most prominent structural difference causing a reduced size of the substrate-binding groove is the deletion of a 13-residue loop in between the two putatively catalytic glutamates. The importance of these latter two residues for catalysis was confirmed by a site-directed mutagenesis study.